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## Nucleosides, Nucleotides and Nucleic Acids

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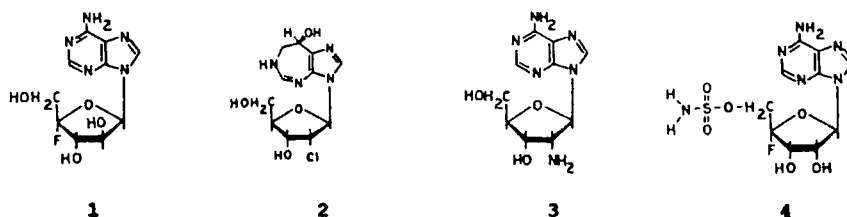
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# BIOSYNTHESIS OF THE NATURALLY OCCURRING NUCLEOSIDE ANTIBIOTICS FROM ADENOSINE

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9- $\beta$ -D-Arabinofuranosyladenine (ara-A, 1), first isolated from the culture filtrates of *Streptomyces antibioticus*, has a broad spectrum of activity against DNA viruses in cell culture and is successfully used in therapy of herpes simplex encephalitis, neonatal herpes, herpes zoster and chronic myelogenous leukemia<sup>1</sup>. 2'-Chlorodeoxycoformycin (2'CldCF, 2), 2'-amino-2'-deoxyadenosine (3) and nucleocidin (4) have been isolated from the culture medium of *Actinomadura* and *S. clavus*, respectively.



[U-<sup>14</sup>C]Adenosine is the direct carbon-nitrogen precursor for these four naturally occurring nucleoside antibiotics. These data were obtained by the addition of [U-<sup>14</sup>C]adenosine to nucleoside antibiotic-producing cultures followed by isolation, crystallization to constant specific activity and hydrolysis to their respective aglycones and pentofuranosyl moieties<sup>2-4</sup>. The ratios of <sup>14</sup>C in the aglycone: pentofuranosyl moieties of ara-A, 2'CldCF, 2'-amino-2'-deoxyadenosine and nucleocidin are summarized in Table 1. Additional proof that the N-ribosyl bond of the [U-<sup>14</sup>C]adenosine is not hydrolyzed during uptake followed by an intracellular resynthesis to form AMP was obtained by the addition of unlabeled adenine simultaneously with the [U-<sup>14</sup>C]adenosine to the

Table 1. Ratios of  $^{14}\text{C}$  in Ara-A, 2'CldCF, 2'-Amino-2'-deoxyadenosine and Nucleocidin

Nucleoside	% <sup>a,b,c</sup>
Ara-A isolated from [2'- $^{18}\text{O}$ ]- and [U- $^{14}\text{C}$ ]adenosine:	
adenine	56
<u>D</u> -arabinose	44
Ara-A isolated from [3'- $^{18}\text{O}$ ]- and [U- $^{14}\text{C}$ ]adenosine:	
adenine	58
<u>D</u> -arabinose	42
2'-Chloro-2'-deoxycoformycin isolated from [U- $^{14}\text{C}$ ]-adenosine:	
aglycone	47
2'-chloro-2-deoxyribose	53
2'-Amino-2'-deoxyadenosine isolated from [U- $^{14}\text{C}$ ]-adenosine:	
adenine	46
2-amino-2-deoxy- <u>D</u> -ribofuranose	54
Nucleocidin isolated from [U- $^{14}\text{C}$ ]adenosine:	
adenine	49
4-fluoro-5-sulfamoyl- <u>D</u> -ribofuranose	51

<sup>a</sup>Ratios of  $^{14}\text{C}$  in the adenosine added to the cultures for ara-A, 2'CldCF, 2'-amino-2'-deoxyadenosine and nucleocidin experiments were: 51:49, 48:52, 48:52, and 54:46, respectively.

<sup>b</sup>For the determination of the  $^{14}\text{C}$  ratios in the aglycone: pentose moieties of the ara-A, 2'CldCF, 2'-amino-2'-deoxyadenosine and nucleocidin, >5000 dpm of crystalline nucleoside was hydrolyzed; recoveries were 81-95%.

<sup>c</sup>The  $^{14}\text{C}$  ratios of the adenosine isolated from the RNA of the organism producing ara-A, 2'CldCF, 2'-amino-2'-deoxyadenosine, and nucleocidin were 57:43, 42:58, 42:58, and 42:58, respectively. Similar ratios of  $^{14}\text{C}$  in the aglycone:pentose were obtained in experiments in which unlabeled adenine was added together with the [U- $^{14}\text{C}$ ]adenosine.

cultures. Isolation of the four nucleoside antibiotics from this experiment showed that the ratios of the  $^{14}\text{C}$  in the aglycone:pentose moieties were the same. Further, the  $^{14}\text{C}$  ratios in the adenine:ribose of the adenosine isolated from the RNA were essentially the same as the  $[\text{U-}^{14}\text{C}]$ adenosine added to the nucleoside antibiotic-producing cultures (Table 1, footnote c).

To study the chemistry at C-2' and C-3' of adenosine during the biosynthesis of ara-A by *S. antibioticus*,  $^{18}\text{O}$ : $^{14}\text{C}$  double label *in vivo* experiments were performed with *S. antibioticus*. The  $[2'\text{-}^{18}\text{O}]$ adenosine and  $[3'\text{-}^{18}\text{O}]$ adenosine were synthesized by Baker and coworkers<sup>5</sup>. In double label *in vivo* experiments with  $[2'\text{-}^{18}\text{O}]$ adenosine (50%  $^{18}\text{O}$  enrichment) and  $[\text{U-}^{14}\text{C}]$ adenosine or  $[3'\text{-}^{18}\text{O}]$ adenosine and  $[\text{U-}^{14}\text{C}]$ adenosine, the  $^{18}\text{O}$ : $^{14}\text{C}$  ratios in the ara-A and adenosine isolated from the RNA were 2.42 and 2.94, respectively. This compares with  $^{18}\text{O}$ : $^{14}\text{C}$  ratios of 2.37 of the  $[2'\text{-}^{18}\text{O}]$ - and  $[\text{U-}^{14}\text{C}]$ adenosine added to the cultures. However, in experiments with  $[3'\text{-}^{18}\text{O}]$  (48%  $^{18}\text{O}$  enrichment) and  $[\text{U-}^{14}\text{C}]$ adenosine, the  $[\text{U-}^{14}\text{C}]$ adenosine was converted to ara-A but there was no detectable  $^{18}\text{O}$  at C-3' of ara-A. The  $^{18}\text{O}$ : $^{14}\text{C}$  ratios for the double label  $[2'\text{-}^{18}\text{O}]$ adenosine and  $[\text{U-}^{14}\text{C}]$ adenosine and the  $[3'\text{-}^{18}\text{O}]$ adenosine and  $[\text{U-}^{14}\text{C}]$ adenosine of the adenosine from the RNA were 2.94 and 3.11, respectively. An enzyme has also been isolated and partially purified from *S. antibioticus* that catalyzes the conversion of adenosine but not AMP, ADP, ATP, inosine, guanosine nor *D*-ribose to ara-A. With the partially purified enzyme, with  $[2'\text{-}^3\text{H}]$ adenosine, the C-2'-H bond was cleaved as evidenced by the release of 8.9% of the tritium from C-2' as tritium oxide. The enzyme displays saturation kinetics, a pH optimum of 6.8, a  $K_m$  of  $8 \times 10^{-4}$  M and is inhibited by heavy metals. Tubercidin, but not sangivamycin, is converted to ara-tubercidin by the enzyme.

With respect to the direct conversion of adenosine (or adenosine nucleotide) to 2'CldCF and 2'-amino-2'-deoxycoformycin, the data show that the hydroxyl at C-2' of the ribosyl moiety of adenosine undergoes a replacement by a chloro or an amino group. Furthermore,  $^{36}\text{Cl}$ chloride when added to the culture medium resulted in the isolation of  $^{36}\text{Cl}$ 2'CldCF. Finally, the direct conversion of  $[\text{U-}^{14}\text{C}]$ adenosine to nucleocidin (4) by *S. clavus* (Table 1) demonstrates that the C-4'-H is stereochemically replaced by fluoride with retention of configuration. The mechanisms for the regioselective modification of the C-2' hydroxyl group and the hydrogen at C-4' of adenosine and the stereospecific insertion of the amino-, chloro-, and fluoro groups are under investigation.

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